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Isolation and Identification of Lathycarpin, a New Pterocarpan Phytoalexin from Lathyrus sativus

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After treatment with the fungus *Helminthosporium car-bonum* or aqueous copper sulphate, excised *Lathyrus sati-vus* leaflets produce pisatin and a second isoflavonoid phytoalexin (lathycarpin) identified as (+)-6aR; 11aR-2,3-di-methoxy-6a-hydroxy-8,9-methylenedioxypterocarpan.

Introduction

Earlier studies [1, 2] have demonstrated that a number of isoflavonoid (pterocarpan) phytoalexins [3] accumulate in excised, fungus-inoculated leaflets of species belonging to the genus *Lathyrus* (Leguminosae-Papilionoideae; tribe Vicieae). Apart from pisatin (3-methoxy-6a-hydroxy-8,9-methylenedioxypterocarpan, 1), the most commonly encountered *Lathyrus* phytoalexin, small quantities of three other fungitoxic pterocarpans (medicarpin, maackiain and variabilin) are variously produced by several *Lathyrus* spp. [2]. In addition, nissolin and its 9-O-methyl ether (methylnissolin) are formed, to-

$$H_3^{CO}$$
 $\frac{3}{1}$ $\frac{4}{1}$ $\frac{6a}{0}$ $\frac{6}{10}$ $\frac{6}{10}$ $\frac{7}{9}$ $\frac{8}{0}$ $\frac{1}{10}$ $\frac{$

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gether with medicarpin, by fungus-treated phyllodes of L. nissolia although as yet these two unusual 3,9,10-trisubstituted pterocarpans have not been discovered elsewhere in the genus [4]. Pisatin and the non-isoflavonoid chromones, lathodoratin methyl-lathodoratin, occur as phytoalexins in tissues of L. odoratus (sweet pea) [5], whilst some evidence has been obtained to suggest that the Indian pulse L. sativus (grass pea; chickling pea) produces 1, and a new pterocarpan (designated LS-2) of undetermined structure [1]. We have now re-examined the phytoalexin response of L. sativus and can confirm the ability of detached leaflets to accumulate 1 following treatment with a spore suspension of Helminthosporium carbonum [1, 2]. This fungus also stimulates the formation of a second antifungal isoflavonoid, probably identical with LS-2, which we propose to name lathycarpin. The characterisation of lathycarpin (+)-6aR; 11aR-2,3-dimethoxy-6a-hydroxy-8,9methylenedioxypterocarpan (2) is described in this report.

Results and Discussion

Pisatin and lathycarpin were initially isolated from H. carbonum-inoculated L. sativus leaflets (but not from those treated with de-ionised H₂O) using the drop-diffusate technique outlined in the Experimental section. This procedure was very tedious, however, and during later stages of the project was replaced by a modified diffusion method (see Experimental) in which leaflets were floated for 10-12days on aqueous CuSO₄. After this period, the CuSO₄ solution was shaken with EtOAc to remove all isoflavonoid compounds, subsequent Si gel TLC purification of the organic phase yielding both 1 and 2 in milligram quantities. Pisatin was identified by UV, MS and Si gel TLC comparison with authentic material obtained from Pisum sativum [6]. Other compounds currently recognised as Lathyrus phyto alexins (see Introduction) were not produced by L. sativus.

The UV (EtOH) spectrum of chromatographically pure lathycarpin (λ max: 212, 235 sh, 303 nm) closely resembled that of 2,3-dimethoxy-8,9-methylenedioxypterocarpan (λ max: 212, 235 sh, 304 nm), and could be distinguished from those of spectroscopically similar 3,4-dioxygenated pterocarpans (e.g. 4-methoxymaackiain) by the absence of two



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slight shoulders between 270 and 290 nm [7]. Aqueous NaOH had no effect on the neutral UV spectrum of lathycarpin, whereas addition of conc. HCl resulted in rapid (1 min) dehydration to afford the corresponding pterocarpene (λ max: 353 and 368 nm). This acid-mediated spectroscopic change is characteristic of pterocarpans (e.g. pisatin and variabilin) containing a labile C-6a OH group [7, 8]. Further evidence for tertiary hydroxylation of 2 (High Resolution MS, M+ 344.0892; $C_{18}H_{16}O_{7}$; 55%) was provided by the mass spectrum which exhibited a major fragment at m/z 326.0803 (M⁺-H₂O; 100%) corresponding to $C_{18}H_{14}O_{6}$ (cf. 1, M⁺ 314 (28%), m/z296 (M⁺-H₂O; 100%)). Other prominent ions in the MS of 2 appeared at m/z 345 (M⁺ + 1; 11%), 329 $(M^+-CH_3; 5\%), 327 (22\%), 325 (31\%), 316 (16\%),$ 311 (16%), 301 (6%), 285 (13%), 283 (7%), 199 (9%), 163 (26%), 151 (9%), 149 (14%) and 133 (10%). On TLC plates sprayed with chromotropic acid reagent, lathycarpin gave a purple-pink colour indicative of methylenedioxy substitution [9, 10].

In view of its co-occurrence with 1, the above data suggested that lathycarpin was a methoxy-pisatin, and this was confirmed by ¹H NMR spectroscopy (Table I) which revealed that pisatin and lathycarpin differed only in that the latter compound possessed an extra methoxyl substituent, and lacked a C-2 proton. The new phytoalexin is thus 2-methoxypisa-

Table I. 1H NMR data for pisatin and lathycarpin a.

Pisatin (1)	Lathycarpin (2)
7.36 d, 1H (<i>J</i> = 8.5 Hz)	6.96 s, 1H
$6.63 \mathrm{q}, 1 \mathrm{H}$ ($J = 8.5, 2.5 \mathrm{Hz}$)	_
$6.39 \mathrm{d}, 1 \mathrm{H}$ ($J = 2.5 \mathrm{Hz}$)	6.43 s, 1H
4.11 s, 2H	$4.06 \mathrm{d}, 2\mathrm{H}$ ($J = 1.5 \mathrm{Hz}$)
6.89 s, 1H	6.89 s, 1H
6.35 s, 1H	6.36 s, 1H
5.29 s, 1H	5.26 s, 1H
$5.93 \mathrm{q}, 2\mathrm{H}$ ($J = 2.9, 0.9 \mathrm{Hz}$)	$5.92 \mathrm{q}, 2\mathrm{H}$ ($J = 2.9, 0.9 \mathrm{Hz}$)
3.75 s, 3H	{ 3.76 s, 3H 3.7 8s, 3H
	7.36 d, 1H (J = 8.5 Hz) 6.63 q, 1H (J = 8.5, 2.5 Hz) 6.39 d, 1H (J = 2.5 Hz) 4.11 s, 2H 6.89 s, 1H 6.35 s, 1H 5.29 s, 1H 5.93 q, 2H (J = 2.9, 0.9 Hz)

^a Solvent, $(CD_3)_2CO$; chemical shifts are given as δ values (TMS reference); figures in parentheses refer to coupling constants.

tin (2,3-dimethoxy-6a-hydroxy-8,9-methylenedioxy-pterocarpan, 2), a structure supported by the loss, relative to pisatin, of significant coupling in the H-1 and H-4 signals and by the upfield shift of H-1 consequent upon *ortho*-oxygenation [11].

Lathycarpin has $\left[\alpha\right]_{589\,\mathrm{nm}}^{21} + 232\,^{\circ}$ (1.2 mg in 1 ml MeOH) and can therefore be assigned the 6aR; 11aR absolute configuration depicted in **2** [12]. Pisatin from *L. sativus* is also dextrorotatory, $\left[\alpha\right]_{589\,\mathrm{nm}}^{21} + 292\,^{\circ}$ (1.05 mg in 1 ml MeOH).

In TLC plate bioassays [13, 14], lathycarpin (20 µg) was clearly inhibitory to the spore germination/germ tube growth of Cladosporium herbarum. More precise measurements against radial mycelial growth of H. carbonum [14] gave an ED₅₀ value of about 45 µg/ml, comparable with that reported earlier for pisatin [15]. On average, diffusates from excised L. sativus leaflets treated with a spore suspension of H. carbonum contained pterocarpans 1 and 2 at concentrations of 18 and approx. 30 µg/ml (based on $\varepsilon = 7244$ at 309 nm for 1 [8]) respectively. Abiotic induction using droplets of aqueous CuSO₄ was less efficient yielding (over four experiments) only $5-14 \mu g/ml$ of 1, and $8-20 \mu g/ml$ of 2. Leaf diffusates invariably contained more lathycarpin than pisatin.

Experimental

Plant material. Seeds of an unnamed variety of Lathyrus sativus L. (supplied by Dr. L. J. G. Van der Maesen, I.C.R.I.S.A.T., Hyderabad, India) were sown in John Innes No. 1 compost, and the resulting plants grown $(20-24\,^{\circ}\text{C})$ for about 6 weeks before individual leaflets were removed for treatment with either H. carbonum or aqueous CuSO_4 . At later stages of growth, the plants were routinely deflowered to encourage leaf production.

Isolation and purification of pisatin (1) and lathy-carpin (2). a) Standard drop-diffusate technique. 2-5 droplets of aqueous CuSO₄ (0.25 g/100 ml de-ionised H₂O, plus 0.5 ml Tween 20 as a wetting agent) or H. carbonum spore suspensions (also containing Tween 20) [16, 17] were placed along the upper surface of excised L. sativus leaflets floating on tap H₂O. After incubation (20 °C) for 72 h, the droplets (diffusate) were collected, extracted (× 3) with EtOAc, and the pooled organic fractions reduced to dryness (in vacuo, 40 °C). Si gel TLC (Merck, F-254,

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layer thickness 0.25 mm) of the extracts (CHCl₃ -MeOH, 50:1, 12 h equilibration at 20 °C) gave pisatin + lathycarpin as a broad, fluorescence-quenching band $(R_F 0.67)$. This band was removed and the pterocarpans eluted (EtOH) prior to separation by Si gel TLC in n-hexane-acetone, 2:1, 1 h equilibration [1] (1, R_F 0.37; 2, R_F 0.28). Final TLC purification of pisatin and lathycarpin was undertaken using n-pentane-Et₂O-glacial HOAc, 75:25:3, 1 h equilibration $(1, R_F 0.31; 2, R_F 0.10)$. b) Modified diffusion technique. In a typical experiment about 600 L. sativus leaflets were cut into short (1-3 cm) sections and floated (10-12 days) on aqueous CuSO₄ (about 21; see above for composition) in transparent-plastic sandwich boxes covered with clear, cling-film food wrap. The leaf material was ultimately discarded, and the $CuSO_4$ solution shaken (×2) with equal volumes of EtOAc. Si gel TLC of the organic phase was undertaken as described above. Pterocarpan yields ranged from 0.5-1.1 mg for pisatin, and from approx. 1.2-1.9 mg for lathycarpin.

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